Impact of Fluoridated Waters on the Density of Mosquito Larvae in the Municipality of Dassa-Zoumé in Benin

Kitidjo Victor Jacques SOSSOU1, Waris Kéwouyémé CHOUTI1-2*, Luc DJOGBENOU3, Romaric AKOTON3, Carine Nelly KELOME3

1Laboratory of Applied Hydrology, National Institute of Water (INE), University of Abomey-Calavi 01 BP: 526 Cotonou, Benin
2Laboratory of Inorganic Chemistry and the Environment, Faculty of Science and Technology (FAST), University of Abomey-Calavi BP: 4521 Cotonou, Benin
3Tropical Infectious Diseases Research Centre (TIDRC), University of Abomey-Calavi, 01BP 526 Cotonou, Benin
*Corresponding author: warichouti@fast.uac.bj

Received June 02, 2022; Revised July 07, 2022; Accepted July 15, 2022

Abstract Mosquitoes are sources of nuisance and vectors of pathogenic agents for humans and animals. The fight against these insects requires a very in-depth knowledge of the ecology of their places of development. This is the reason why a study on the impact of fluoride on the development of mosquitoes was carried out in the municipality of Dassa-Zoumé, in central Benin. Thus, larval habitat has been actively sought and prospected. The physicochemical and chemical (e.g.: fluorine) parameters of the water in these development sites were measured. A total of 38 larval habitats were identified and surveyed. Most larval habitats are in the immediate environment of human populations. The characterization of the substrate at the level of these development sites has informed us about the ecological requirements of larval density. Indeed, the analysis of the results obtained shows that the individual increase in fluoride ions, salinity, turbidity, suspended matter or exposure to the sun of the site favors the increase in the number of deposits of development collected. On the other hand, the opposite effect is obtained with increasing chlorides ions or hydrogen potential. In addition, the combined action of fluoride ions and a few other elements inhibits the density of larvae in the stations. These results show that to limit or inhibit the density of larvae in this region, the presence of fluoride ions combined with either a high concentration of chloride ions and / or a high amount of salinity is required.

Keywords: fluorine, fluoride, larval habitat, mosquitoes, Dassa-Zoumé


1. Introduction

Malaria infections are a major public health problem and the main cause of morbidity and mortality, particularly among children under five and pregnant women. A global estimate of 229 million new malaria cases and around 409,000 malaria deaths were reported in 2019, with 92% of the disease burden recorded in the African regions [1]. Malaria is transmitted through the bite of female Anopheles mosquitoes, which carry the infection by protozoan parasites Plasmodium species to humans [2,3].

In most sub-Saharan African regions, the main malaria vectors are An. gambiae s.l. An. funestus s.l. An. nili and An. moucheti. In Benin, An. gambiae s.l. remains the major malaria vector, with variable vectorial capacities and behaviors [4]. Immature An. gambiae s.l. develops in a variety of aquatic ecosystems, such as fresh, brackish waters found in rural, coastal, and urban areas, where larvae emerge into adult mosquitoes. Water quality of larval habitat is an important determinant in egg laying, for adequate growth and development from larval stages until adults [5,6,7,8]. An. gambiae, breeds in natural/artificial permanent and semi-permanent water bodies with floating or emerging vegetation like the edges of swamps, in weedy and grassy parts of rivers, streams, furrows, ditches and ponds with low salinity and little richness in organic matter [9]. It was reported that larval habitat characteristics could influence several life-history traits in mosquitoes, such as oviposition preferences, hatching, immature development, pupation [10]. Consequently, the habitat characteristics, especially physicochemical, could influence adult productivity, and it will have a significant impact on malaria transmission.
In Benin, despite the efforts made by the authorities in charge of health for many years, malaria remains a real public health problem in most localities with an increase in the incidence of malaria and, in cases of fatalities, which has been multiplied by two between 2010 and 2017 [11]. The short reviews on the effect of physics (pH, total suspended solids, total dissolved solids, electrical conductivity, etc.) and mostly fluorine on larval density and development must be provided here with interest to the vector control strategies.

In addition, since insecticide resistance is common in Anopheles mosquitoes and the existence of residual transmission when people are not protected by vector control tools (Indoor Residual Spray or Long-Lasting Insecticidal Nets), the interest in developing complementary larval vector control methods is important. Information on the physicochemical properties of the larval habitat, a potential key element for larval surveillance, could help in the implementation of better vector control programs. Concomitantly with the results of this research, one of the vector control methods recommended by the National Malaria Control Program (PNLP) is based on the use of insecticides. Indeed, a study carried out on mosquitoes has revealed that the insecticides used to fight against mosquitoes and containing sulfuryl fluoride as an active substance are more effective [12]. It should be deduced from these results that areas with water resources characterized by the presence of fluorine can reduce the proliferation of anopheles breeding grounds. Although, many studies have been carried out in Africa on the malaria vector bioecology, none of them have establish a link between the presence of fluorine in Anopheles larval habitats and their abundance their density.

Since relatively little information is available on larval habitat containing fluorine and the abundance of An. gambiae mosquitoes in the field settings, the present study aimed to find the relationship between this physicochemical characteristic with the abundance of An. gambiae s.l., in the municipality of Dassa-Zoumè in Benin, for effective vector monitoring and disease control strategy.

2. Methods
2.1. Study Site and Sample Collection
Potential *An. gambiae* s.l. larval habitats were randomly selected during each season where sampling was made. Larvae collections were carried out in August 2020 (dry season) and in October 2020 (rainy season). Nine (09) and twenty-nine (29) potential *An. gambiae* s.l. larval habitats were prospected (Figure 1). Mosquito larvae were collected using dipping method [13]. In each plot, 20 scoops were taken using a standard white 300 ml dipper (ladle). Mosquito larvae collected were identified morphologically using identification keys [9]. The physical parameters such as habitat positivity for *Anopheles* larvae (presence or absence of *Anopheles* larvae in the habitat), larvae count (instar stade count of *Anopheles* larvae), sun/shade (whether the water in the habitat is exposed to direct sun or it is shaded by the presence of vegetation or any other covering), were recorded on site by visual observation.

To assess the physicochemical characteristics of prospected larval habitat, water samples were aseptically collected in cleaned glass bottles boxes of 0.5 liters to avoid interference before immersing it in water. The samples were stored in coolers fitted with frozen accumulator at a temperature between 2°C and 8°C during transportation to the laboratory. Water collections in which the pre-imaginal mosquito stages were considered as larval habitat. Each larval habitat was georeferenced using a GARMIN® brand GPS and the data imported into Arc GIS version 10.1 mapping software for mapping.

### 2.2. Physicochemical Parameters of Breeding Water

The samples are analyzed at the water and food quality control laboratory of the National Directorate of Public Health of Benin by standardized methods and techniques. The hydrogen potential (pH) is measured in water samples using a WTW 340i pH meter. Conductivity was measured by applying alternating electric current to two electrodes immersed in a solution and measuring the resulting voltage. The differential weighing method was used for the measurement of Suspended Solids (SSD). Chloride ions are determined by the MOHR method. The analytical method used for the determination of fluoride is the SPADNS colorimetric method, based on a HACH DR/2010 spectrophotometer. Iron was not determined, but compensatory measures were taken prior to the determination of fluorine, adding to each sample a drop of tri-ethanolamine intended to complex the iron, thus preventing it from creating interference.

### 2.3. Data Analysis

Excel and Rstudio software were used for data processing and graphical visualization. In addition, to highlight the links between the variables, the generalized linear model (GLM) was used. Since the target variable **Y** corresponding to larval density is a counting variable, Poisson regression was used as a prediction model. Thus, Poisson regression was used to establish, on the one hand, the link between fluoride content and larval density and, on the other hand, between the interaction of fluoride with each of the other parameters studied and larval density. The Stata 15 software was used to establish generalized linear models as part of this research.

### 3. Results and Discussion

#### 3.1. Results

##### 3.1.1. Typology of Larval Habitats

The types of larval development sites are presented in Table 1. The table shows that 77.78% of the deposits prospected are artificial and 22.22% are natural in dry season whereas 100% of sites are artificial in wet season. Also, 88.88% of the lodgings are sunny and 11.12% are shaded in dry season respectively compared to 62.07% and 37.93% in wet season.

<table>
<thead>
<tr>
<th>Site types</th>
<th>Sun exposure</th>
</tr>
</thead>
<tbody>
<tr>
<td>Natural</td>
<td>Artificial</td>
</tr>
<tr>
<td>Dry season</td>
<td>22%</td>
</tr>
<tr>
<td>Wet season</td>
<td>0%</td>
</tr>
</tbody>
</table>

##### 3.1.2. Global Visualization of Parameters

The spatial distribution of the physico-chemical characteristics (logarithmic scale) and the larval density (arithmetic scale) of the waters of the deposits in the dry and wet season are presented respectively in Figure 2 and Figure 3.
3.1.3. Physicochemical Characterization and Larval Density in Dry Season

The boxplots show the dispersion and the centralization of physico-chemicals parameters and larval density of sites in dry season.

The distribution of values shown in Figure 4 shows that the larval density is high when the fluoride concentration is low (<0.05 mg/L) in dry season. The values of this boxplot are evenly distributed around the median (150 larvae).

![Figure 4. Distribution of fluoride (F) and larval density (Densite) in dry season](image)

![Figure 5. Distribution of Chloride (Cl), Salinity (Sal), Suspended Solids (MES), Electrical Conductivity (CE), Total Dissolved Solids (TDS), Turbidity (Turb) and larval density (Densite) in dry season](image)
There is a similar distribution of graphs between Chloride and Salinity, Suspended Solids and Turbidity, Electrical Conductivity and total dissolved solids (Figure 5). As opposed to fluorides, the larval density is high when the values of these parameters are high in dry season.

3.1.4. Physicochemical Characterization and Larval Density in Wet Season

Are presented below the boxplots of physico-chemicals parameters and larval density of sites in wet season.

The distribution of values (Figure 6) shows that the median value of larval density is high in three points. The two highest values correspond to a relatively high fluoride concentration.

According to Figure 7, the high larval densities obtained have a high hydrogen potential. The high pH in question is around 6.8 and 7.5. Depending on the class, they tend from neutrality to basicity.

The analysis of Figure 8 does not allow us to apparently show a correlation between the different parameters. We will therefore move on to statistical analysis to better interpret the data.
3.1.5. Impact of Physico-chemical Parameters on Larval Density

Table 2 presents the successive results of the various Generalized Linear Models (MLG) according to Poisson regression.

According to the table above the probability Pr(>|z|) is less than 5% in virtually all cases except for the chloride variable of the MLG 2 model and fluoride of the MLG 9 model. This demonstrates that the other models are significant and can be used.

- MLG 1: The logarithm of the mean number of larval densities increases by 1.34 as the amount of fluoride increases by one unit in mg / L. That is, the larval density when fluoride increases by one unit in mg / L is 3.82 (i.e.: $e^{1.34}$) times greater than the starting (previous) larval density. Thus, fluorides alone promote larval development.
- MLG 2: The larval density when the simultaneous fluoride * chloride interaction increases by one unit is 0.23 (i.e.: $e^{-1.27}$) times lower than the starting (previous) larval density. We can say with reservation from further testing that the combination of fluoride * chloride has an inhibitory influence on mosquito larvae.
- MLG 3: The larval density when the simultaneous interaction fluorides * salinity increases by one unit is $1.14 \times 10^{-8}$ (i.e.: $e^{-10.23}$) times lower than the starting (previous) larval density. We can say with reservation from further testing that the association of fluoride * salinity has a considerable inhibitory influence on mosquito larvae.
- MLG 4: The larval density when the simultaneous interaction of fluorides * electrical conductivity increases by one unit is 0.997 (i.e.: $e^{0.0031}$) times lower than the starting (previous) larval density.

Given that $0.997 \approx 1$ we can deduce that the influence of the association fluoride * electrical conductivity on the larvae is negligible.

- MLG 5: The larval density when the simultaneous interaction fluorides * turbidity increases by one unit is 0.985 (i.e.: $e^{0.005}$) times lower than the starting (previous) larval density. Given that 0.985 $\approx 1$ it can be deduced that the influence of the fluoride * turbidity association on the larvae is negligible.
- MLG 6: The larval density when the simultaneous interaction fluorides * TDS increases by one unit is 0.9 (i.e.: $e^{-0.1}$) times lower than the starting (previous) larval density. Given that 0.9 $\approx 1$ it can be deduced that the influence of the fluoride * TDS association on the larvae is negligible.
- MLG 7: The larval density when the simultaneous interaction fluorides * SS increases by one unit is 0.99 (i.e.: $e^{-0.006}$) times lower than the starting (previous) larval density. Given that 0.99 $\approx 1$ it can be deduced that the influence of the fluoride * SS association on the larvae is negligible.
- MLG 8: The larval density when the simultaneous interaction of fluorides * pH increases by one unit is 3361 (i.e.: $e^{15.8}$) times greater than the starting (previous) larval density. We can say with reservation from further testing that the combination of fluoride * pH has a considerable favorable influence on mosquito larvae.
- MLG 9: The larval density when the simultaneous interaction fluorides * sun exposure increases by one unit is 15.8 (or $e^{5.5}$) times greater than the starting (previous) larval density. We can say with reservation from further testing that the combination of fluorides * sun exposure has a favorable influence on mosquito larvae.

### Table 2. Generalized Linear Modeling (MLG) / Poisson Regression

| Model | Number of larvae | Estimated value | Standard error | Z statistic | Pr(>|z|) |
|-------|-----------------|----------------|----------------|-------------|----------|
| MLG 1 | Constant        | 2.825447       | 0.0718395      | 39.33       | 0.000*** |
|       | Fluorides       | 1.339357       | 0.1065249      | 12.57       | 0.000*** |
| MLG 2 | Constant        | 2.487408       | 0.2328246      | 10.68       | 0.000*** |
|       | Fluorides       | 4.566047       | 0.3879404      | 11.77       | 0.000*** |
|       | Chlorides       | -0.0155779     | 0.1162371      | -0.13       | 0.893    |
|       | Fluorides * Chloride | -1.474403 | 0.1949865 | -7.56     | 0.000*** |
| MLG 3 | Constant        | 1.352792       | 0.1850875      | 7.31        | 0.000*** |
|       | Fluorides       | 2.849114       | 0.251312       | 11.34       | 0.000*** |
|       | Salinity        | 16.54462       | 1.7091         | 9.68        | 0.000*** |
|       | Fluorides*Salinity | -18.29025 | 2.220854 | -8.24     | 0.000*** |
| MLG 4 | Constant        | 1.374002       | 0.2498894      | 5.5         | 0.000*** |
|       | Fluorides       | 2.214421       | 0.3294819      | 6.72        | 0.000*** |
|       | $\sigma$ ($\mu$S/cm) | 0.0038859 | 0.0005966 | 6.51     | 0.000*** |
|       | Fluorides*$\sigma$ ($\mu$S/cm) | -0.0030793 | 0.0007692 | -4       | 0.000*** |
the larval density of the deposits. Nevertheless, we notice that in the dry season, a high concentration of fluoride does not increase the larval density, while this is the case in the rainy season. On the other hand, it should be noted that overall salinity, total dissolved solids, suspended solids, turbidity and electrical conductivity increase proportionally with larval density, regardless of the season. As for the hydrogen potential (pH), the results clearly show that the increase in larval density is conditioned by high pH values (ranging from neutral to basic). These results agree with those obtained by [17] as well as [18] who showed the impact of the physicochemical characteristics of the larval stations, in particular the hydrogen potential, which seems to play a role in the non-proliferation of the larvae [19]. Indeed, Anopheles larvae prefer well oxygenated waters with basic pH [20]. In other words, the higher the pH increases (basic), the higher is the number of larvae in the breeding places. Likewise, the results obtained concerning the influence of the exposure of sites to the sun are those also obtained by [14] according to which the larvae of anopheles prefer fresh water, exposed to sunlight and at high temperatures. Moreover, according to the work carried out by [21], the females of Anopheles gambiae sl prefer to lay their eggs in sunny water collections and, according to [22] the temperature and salinity can also promote or limit the growth of larvae.

We deduce that the variation in larval density considers several physico-chemical, chemical, and environmental
parameters. However, the generalized linear model allows us to highlight the interaction between the different parameters studied. The combined action of fluoride ions and other parameters (chloride ions, salinity, conductivity, suspended solids or dissolved solids) limits or inhibits the development of larvae in stations. When fluoride ions and pH are all high or fluoride ions are rising, and the station is exposed to the sun, the larvae develop easily.

Our results showed that to limit or inhibit the density of larvae in the region, it is necessary to act mainly on pH (make the environment acidic) and sun exposure (shade larval sites). One can also act on the combined effect of fluoride and chloride, as well as fluoride and salinity. But controlling these effects will be visibly complicated and uncertain.

4. Conclusion

The results of this research reveal a heterogeneity of larval sites in terms of physicochemical and chemical parameters and environmental conditions. These sampling sites are mostly born from anthropogenic activities and are in the immediate environment of the human populations in the study area. In the light of all the results of this study, it appears that the impact of fluorine on larval density does not follow a standard chronology but also depends on other parameters and the environmental context of the environment taken simultaneously. Nevertheless, an apparent correlation between pH, shading and larval density has been demonstrated. Thus, acidic water not exposed to the sun prevents the development of larvae. Given the complexity of aquatic environments, a more appropriate study would be needed to understand the behavior of larval ecosystems to manage them effectively and contribute to the fight against malaria.

Statement of Competing Interests

The authors have no competing interests.

References